
New Developments in BioWin 4.0

BioWin 4.0 continues to expand the capabilities of wastewater treatment plant simulation with key enhancements to the core model, the model library, and the interface. A focus in this release has been to extend and refine the core Activated Sludge Digestion Model (ASDM) model, which now contains 82 processes acting on 46 state variables. In addition to adding 22 new process rate equations, refinements to the core model have resulted in faster dynamic simulations, particularly for nutrient removal systems.

New Models and Processes

The most significant ASDM extension has been to include three mechanisms for modeling nitrous oxide (N_2O) production and emission. As a result, BioWin 4.0 is a powerful tool for evaluating plant-wide greenhouse gas (GHG) emissions of nitrous oxide, methane and CO_2 . The basis of the N_2O model, and other extensions and improvements to the ASDM are outlined further below.

A new element for trickling filters has been added for increased modeling flexibility. Other related biofilm-based units (*e.g.* SAF, BAF) are planned in an upcoming release.

Model Improvements

Improvements to biological models in BioWin 4.0 have extended their flexibility and improved model predictions. These include:

- Modifications to denitrification modeling to accommodate nitrous oxide modeling.
- A process for slow decay of endogenous residue (with a default zero rate).
- Modifications to the PAO model to allow users to apply either the Delft maintenance approach or the decay/lysis approach.
- A unified kinetic expression for hydrolysis of particulate substrate in activated sludge and anaerobic digesters.
- For biofilms, the default EPS factors for nitrifiers (AOBs and NOBs) have been increased. This change was based on extensive assessment of nitrification performance in IFAS systems.
- Stoichiometry to allow specifying different N and P contents in each biomass and in endogenous residue.
- Gas phase modeling has been rationalized, and all gases now use the same form of Henry's Law expression to determine saturation concentrations. These are editable, and the user can specify temperature dependency.
- For bioreactors, upper and lower air flow limits can be set and also manipulated from BW Controller.
- SRT calculations can include or exclude biofilm mass for attached growth systems.

Usability Improvements

BioWin 4.0 usability improvements include:

- Improved display of the model stoichiometry matrix.
- Restructuring of the Help.
- New additions to the cabinet examples.
- Influent elements remember 'low' flow unit settings.
- Comma display for thousand separators.
- Expanded copy/paste/print facilities in state vectors.
- Total N reported in Explorer.

Model Builder usability improvements include:

- An improved layout.
- A cabinet of examples has been added (eg. ASM1, ASM2d, ASM3).
- A recently used files list and a notes section to document your Builder files.
- A simplified procedure for loading and saving .mod and other recognized file formats.

Improvements to the Album have also been made which include:

- Groupings to assist in finding element specific items for tables and charts. Group titles are numbered and each group is sorted alphabetically.
- Improved interpretation of "Mass rates" for element specific variables.
- Tabulation and plotting of model process rates in bar or time series charts.
- Exporting individual chart data to various formats from the right-click menu.
- Enhanced *Chart Master* settings. Specify defaults for current value and time series plots independently as well as default tools. [In *Chart Master* double-click the plot on the *New chart template* tab].
- Formatted charts for reporting nitrous oxide outputs (current values or time series).

The remainder of this document provides more information on the additions and changes in BioWin 4.0. Further details on each item can be found in the manual and Help.

NEW MODELS

New Model – Nitrous Oxide Emission

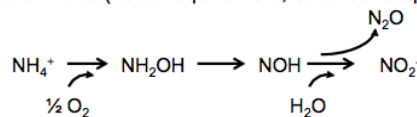
Nitrous oxide has a global warming potential 300 times higher than carbon dioxide and can be a significant source of greenhouse gas (GHG) emissions from wastewater treatment plants. EnviroSim conducted an extensive multi-year study on the GHG issue of nitrous oxide production and emission from activated sludge processes. A comprehensive review of published literature was conducted, assembling state of the art knowledge and resolving incompatibilities amongst the various approaches. Much experimental work was performed to substantiate our understanding and support the development of nitrous oxide models in the BioWin ASDM model.

The outcome of these efforts is embodied in the ASDM model as three major process mechanisms for potential nitrous oxide production. Two of these are mediated by ammonia oxidizing biomass (AOBs) and one by heterotrophs in denitrification, as described below.

1. **Nitrification byproduct:** When AOBs are operating at maximum rate in the presence of ammonia excess, and with no oxygen limitation, a small fraction of the oxidized ammonia is directed to N_2O .

Nitrification by-product

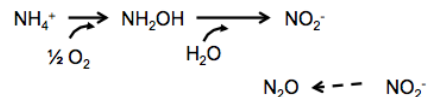
- If AOBs operating at μ_{max} (then NO_2^- present, and sometimes accumulating) then a small portion of oxidized ammonia goes to N_2O
 - Fraction to N_2O proportional to NH_3 processed (i.e. more NO_2^- , more N_2O)
- Occurrence conditions:
 - Significant ammonia ($NH_3 \gg 2 \times K_s$)
 - No oxygen limitation ($DO \gg 0$)
 - Presence of nitrite (not a requirement, but a consequence)



2. **Nitrifier denitrification by AOBs:** Under oxygen-limited conditions where nitrite is present, free nitrous acid (FNA) can be used as a terminal electron acceptor, and is converted to N_2O .

Nitrifier denitrification

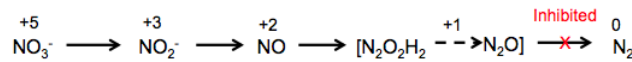
- At low DO, some Nitrifier denitrification
 - The N_2O comes from NO_2^- consumed as TEA (not directly from NH_4^+ or NH_2OH oxidation)
- Occurrence conditions:
 - Low DO $\neq 0$ ($0 < \text{DO} \leq 0.4 \text{ mg/L}$)
 - Active AOBs (generation of NH_2OH)
 - Presence of NO_2^- (FNA) as TEA



3. **Heterotrophic denitrification:** At low DO, and depending on nitrite concentration and pH, free nitrous acid (FNA) reaches a level where the final step of denitrification is inhibited, and N_2O accumulates.

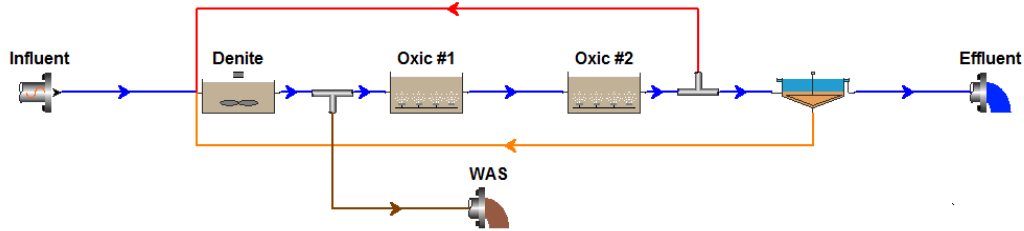
Heterotrophic denitrification

- If $0.4 < DO < 0.8$ (approx.), and NO_2^- and pH such that FNA significant, N_2O may accumulate
- Occurrence conditions:
 - Carbon substrate
 - Presence of nitrate/ nitrite as electron acceptor
 - Inhibiting factors for denitrification (H_2S , 1998_WST_Schonharting)
 - pH: N_2O reductase is sensitive to pH; optimal pH range 7.5 – 8.0
 - **FNA**
 - $DO \neq 0$



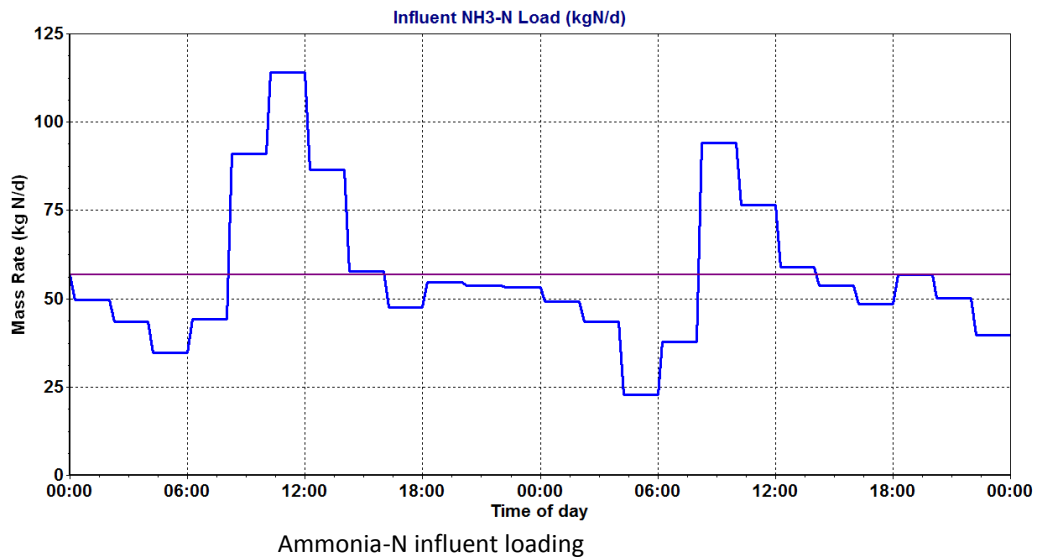
BioWin 4.0 Nitrous Oxide Model Example

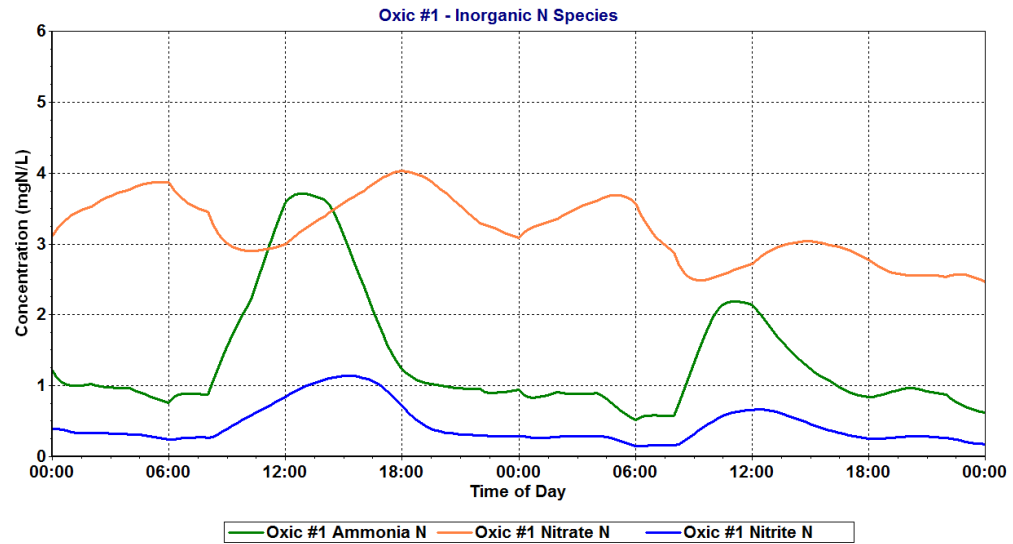
The following example illustrates BioWin 4.0 nitrous oxide modeling capabilities using data from a pilot plant operated by the Swiss Federal Institute of Aquatic Science and Technology (EAWAG). Mixed liquor is recycled from the aerobic zone to an anoxic zone for denitrification at a ratio of 150% of the influent flow. The clarifier underflow is recycled to the anoxic reactor at a ratio of 250% of the influent flow. The SRT is maintained at 12.5 days.



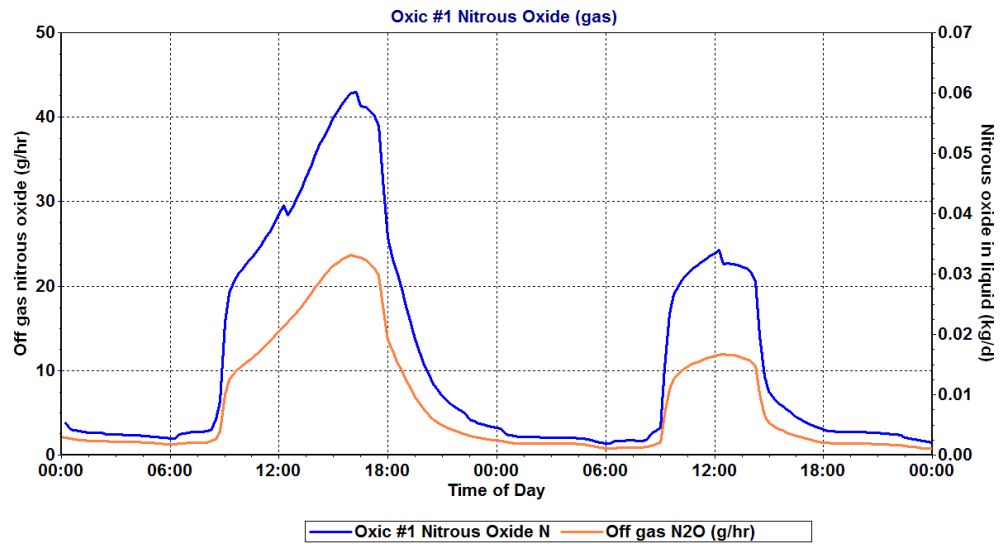
Pilot Plant Process Diagram

The following three plots show the varying influent ammonia-N load over two days, the varying nitrogen species concentrations (ammonia, nitrate, and nitrite) in the first aerated bioreactor [Oxic #1], as well as the mass rate of nitrous oxide in the off-gas and the liquid output from Oxic #1. The ammonia concentration in Oxic #1 increases above 2 mgN/L when the influent ammonia load peaks. Simultaneously the nitrous oxide emissions elevate in the presence of excess ammonia (Mechanism #1).





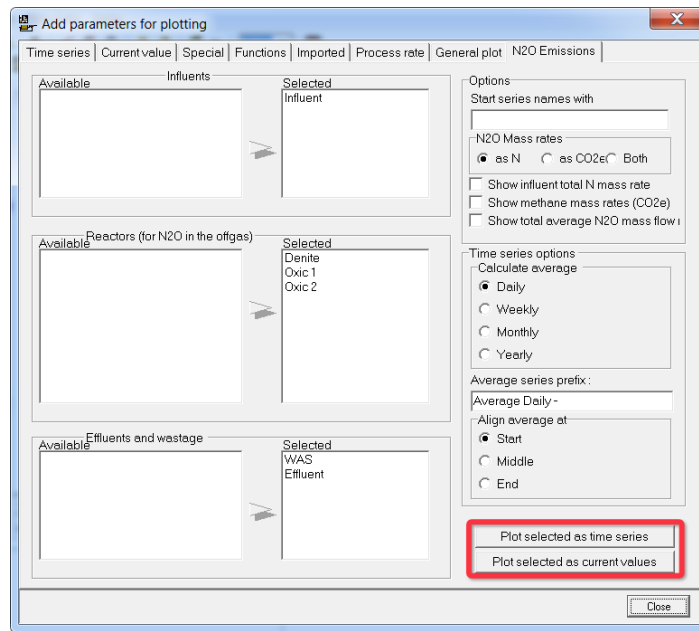
Nitrogen species in aerated bioreactor Oxic #1



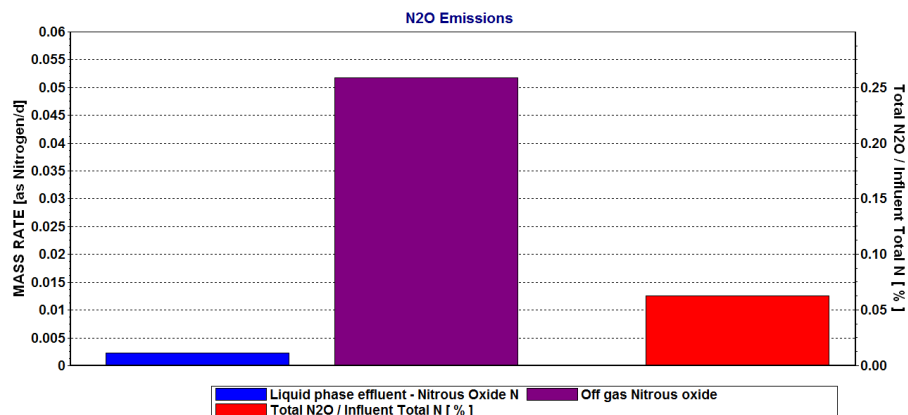
Nitrous Oxide from aerated bioreactor Oxic #1

BioWin 4.0 provides the means for rapidly generating a plant-wide plot of nitrous oxide emissions. To set up the N_2O emissions chart:

- Influent elements are selected to calculate the total nitrogen input load;
- Reactors are selected to estimate off-gas N_2O emission;
- Waste/ effluent streams are selected to estimate N_2O in the liquid stream outputs.

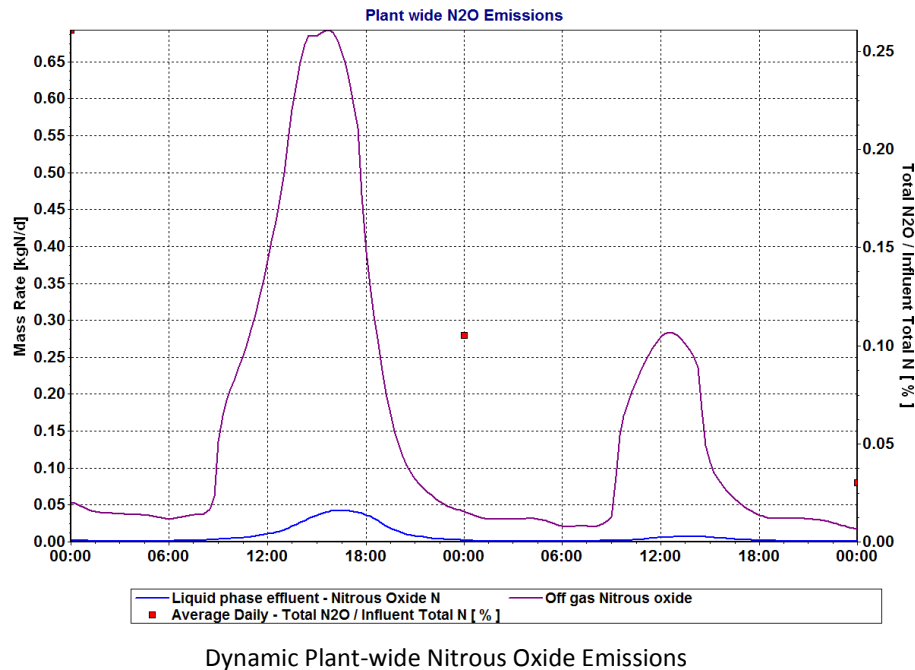


A current value plot typically is selected to demonstrate steady state estimation results. The mass rate of N_2O in the outflows and the total mass rate of N_2O leaving the system via gas and liquid are plotted against the left axis in units of kgN/day (or lbN/day), while the N_2O emission via off-gas as a percentage of the total influent nitrogen load is plotted to the right axis.



Steady-State Plant-wide Nitrous Oxide Emissions

A time series chart should be selected to demonstrate dynamic estimation results. In addition to the mass rate of N_2O in the effluent, total mass rate of N_2O in the off-gas and liquid, and the percentage of N_2O in off-gas out of influent nitrogen, the daily average percentage N_2O emission in off-gas is also plotted.



New Element – Trickling Filter

A new process flowsheet element has been added for Trickling Filters. The TF unit can be configured for various default packing types (eg. rock or plastic media), or the user can customize packing characteristics. The flowsheet image for the trickling filter changes depending on the type of media selected.

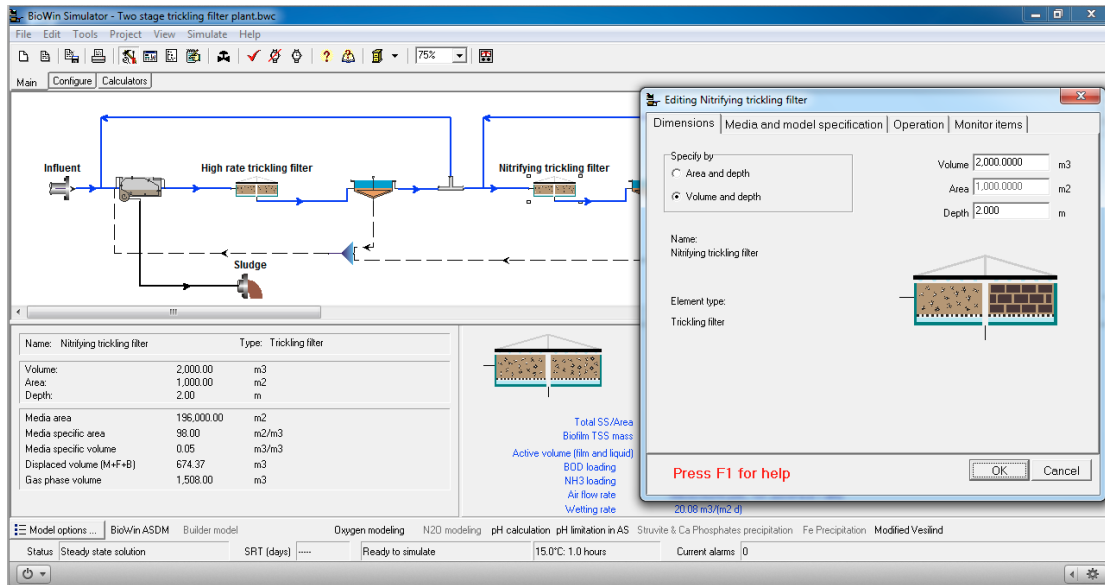
The depth of the trickling filter is divided into three sections or ‘slices’ of equal height with liquid distributed evenly over the top surface. The sections are used for modeling aspects such as oxygen transfer from the top of the trickling filter to the bottom (if the gas phase modeling feature of BioWin is used) and also to simulate removal gradients down the depth of the trickling filter.

On the *Media and model specification* tab the user specifies the type of trickling filter media to be used. Note that trickling filter elements in BioWin are assumed to be 100% full of media. The following media choices are available: Rock, Horizontal, Structured plastic (crossflow), Loose media (random), and Custom. Toggling between the first four of these changes the specific area and specific volume (i.e. 1-porosity) fields to commonly accepted values for those parameters. For custom media, the specific area and specific volume can be adjusted. Users also specify the number of layers to use for modeling the biofilm attached to the media and the liquid boundary layer thickness.

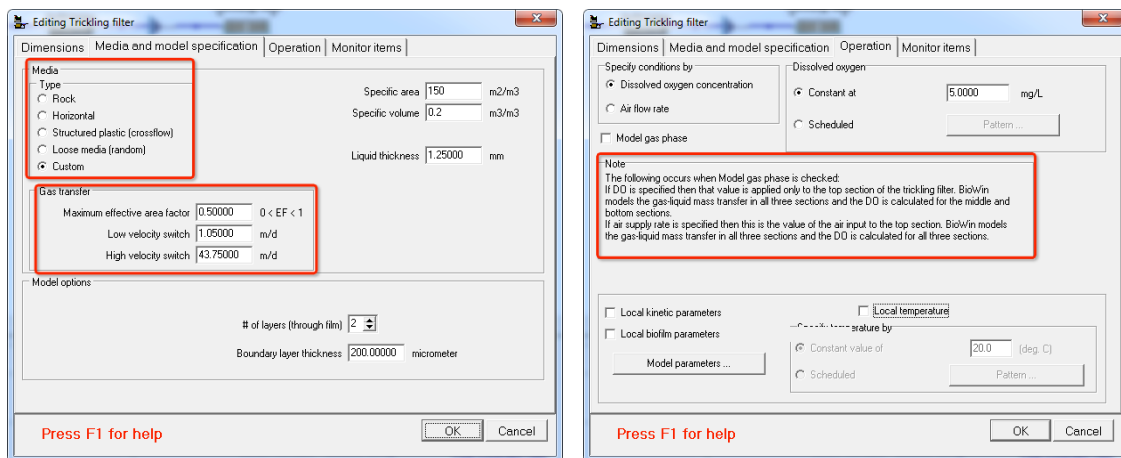
The total thickness of the liquid layer covering the biofilm can be specified for any type of media. This is used to calculate the hydraulic residence time of the trickling filter. Making this parameter larger will increase the hydraulic residence time, and making it smaller will decrease the hydraulic residence time (all other things being equal). Performance of the trickling filter is related to hydraulic loading by making the

area available for gas transfer to the liquid phase dependent on the liquid velocity through the trickling filter (thereby making more or less oxygen available to the biofilm).

Oxygen transfer is a most important factor in determining trickling filter behaviour. The user has considerable flexibility to manipulate aeration through specifying DO concentrations or air flow rates, and selecting whether to model gas phase composition. DO concentrations in the liquid phase can be specified as uniform for each section, or BioWin can model the profile through the filter.



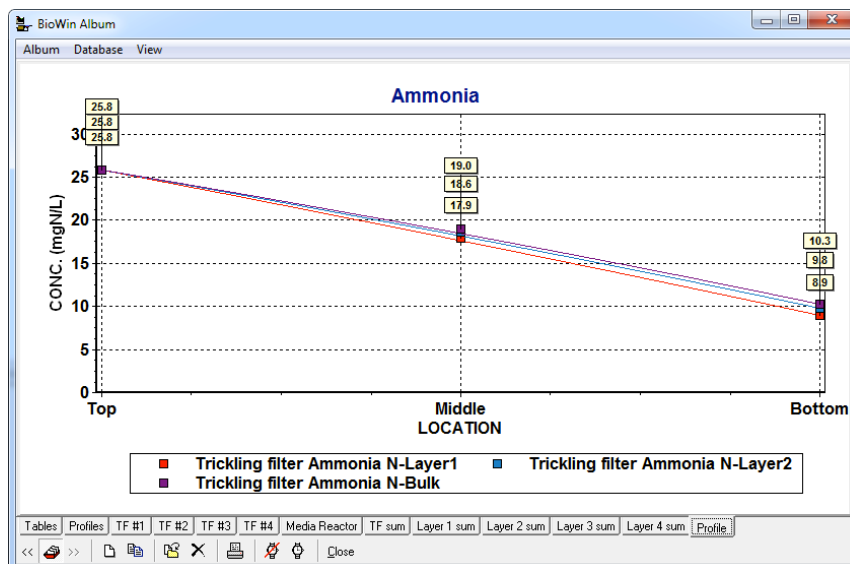
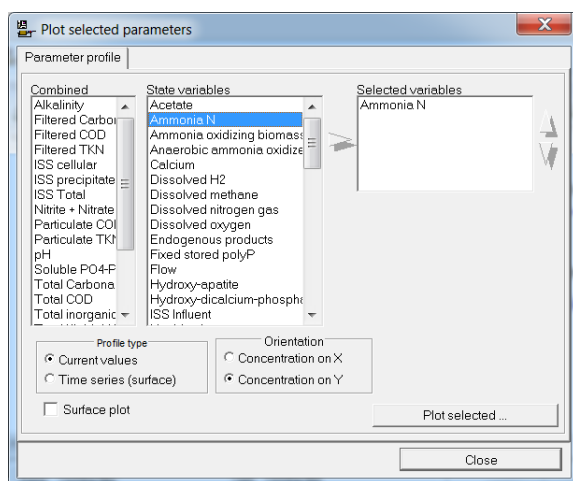
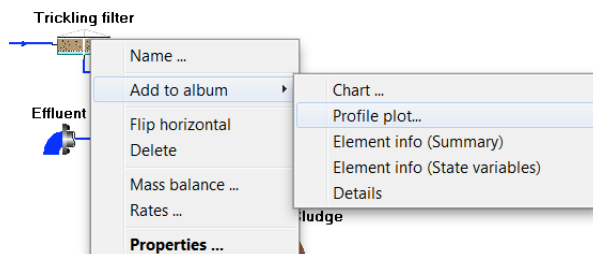
Trickling Filter – Editing Dimensions



Trickling Filter – Media and Model Specification

Trickling Filter – Operating Parameters

Profiles can be plotted to show concentration gradients through the depth of the trickling filter (top, middle, and bottom 'slice') as well as the concentrations in each biofilm layer and in the bulk liquid within each 'slice' of filter.

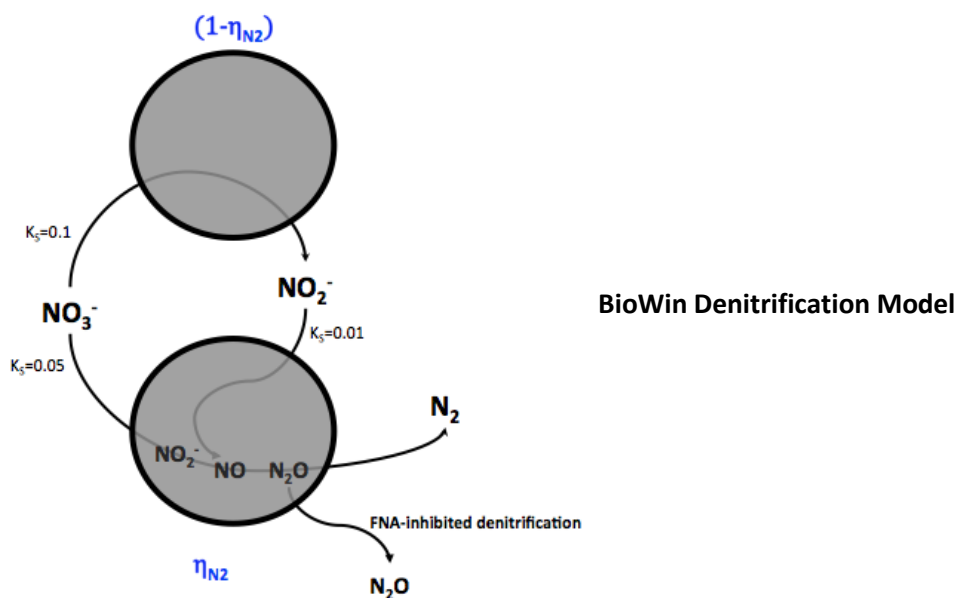


Trickling Filter Ammonia-N Concentration Profile

MODEL UPDATES

Model Modification – Denitrification

EnviroSim's ongoing model development and in-house experimental work has led to the adoption of a novel denitrification model structure. In BioWin 4.0, denitrification follows two pathways within the heterotroph population. One fraction of the population reduces nitrate only as far as nitrite. The remaining population fraction reduces both nitrate and nitrite to nitrogen gas (without releasing intermediates under normal circumstances). Where nitrite is present in the liquid, and free nitrous acid (FNA) concentration is significant (also depends on pH), this population fraction can reduce NO_x to dissolved N₂O rather than nitrogen gas.



In BioWin 3.1, denitrification is modeled as a two-step series process. That is, nitrate is reduced to nitrite (and the nitrite implicitly is excreted to the liquid phase), and then nitrite is taken up and converted to nitrogen gas, without tracking intermediates. In recent modeling literature, particularly where nitrous oxide is a concern, a 4-step denitrification path (nitrate - nitrite - nitric oxide - nitrous oxide - nitrogen gas) is often adopted. In these approaches, it is implied that each intermediate is expressed into the liquid phase and immediately taken up again for the subsequent step; seemingly impractical from an energetic point of view.

The BioWin 4.0 model was implemented after extensive investigation and research which included a comprehensive literature review and experimental work. Experiments conducted in support of denitrification model refinement included unaerated batch tests combining mixed liquor and municipal wastewater for the evaluation of issues such as (a) nitrate *versus* nitrite removal rates when batch tests are spiked with each electron acceptor separately; (b) nitrite accumulation; and (c) removal rates when both nitrate and nitrite are available in similar concentrations.

As a result of this effort, the BioWin 4.0 denitrification model now allows for tracking of key intermediates such as nitrous oxide, and a robust mathematical approach which better approximates the denitrification process.

Model Addition – Decay of Endogenous Residue

BioWin allows for the conversion of endogenous decay products to biodegradable particulate substrate. This is modeled as a first order process with respect to the concentration of the endogenous products. Nitrogen and phosphorus included in the endogenous matter are lysed in parallel as particulate organic N and P. The default process rate constant is set at zero. The user should specify a small value (e.g. 0.007 /day) to simulate this slow decay process.

Model Extension – PAO Lysis Decay *versus* Maintenance Approach

Several studies of biological phosphorus removal in EBPR systems in recent years have focused on systems implementing non-conventional methods such as fermentation of return activated sludge (RAS) or mixed liquor solids (MLSS) as a means for improving EBPR performance. Regarding modeling of EBPR in these non-conventional systems, it has been suggested that the method for modeling PAO biomass decay may be an important factor in correctly predicting EBPR performance [e.g. Houweling *et al.*, 2010]. Specifically it has been suggested that the maintenance approach proposed by the Technical University of Delft (TUD) may offer some advantages over the University of Cape Town decay/lysis approach applied in BioWin.

The ability to choose the Delft approach in place of the BioWin approach has been included in BioWin 4.0. Rate parameters for the maintenance approach are assigned values of zero. The user should assign appropriate values (and set PAO aerobic/anoxic and anaerobic decay rates to zero) to apply the Delft approach if desired.

It is worth noting that both decay approaches have been evaluated recently as part of a WERF study. The general finding was that either approach has little impact on EBPR performance predictions. This likely is due to the slow decay rates of PAOs, and that decay has a limited impact on model predictions compared to other model processes.

Kinetic parameter editor

AOB NOB ANAMMOX OHO Methylootrophs PAO Acetogens Methanogens pH Switches

Parameters

Name	Default	Value	Arrhenius
Max. spec. growth rate [1/d]	0.9500	0.9500	1.0000
Max. spec. growth rate, P-limited [1/d]	0.4200	0.4200	1.0000
Substrate half sat. [mgCOD(PHB)/mgCOD(Zbp)]	0.1000	0.1000	1.0000
Substrate half sat., P-limited [mgCOD(PHB)/mgCOD(Zbp)]	0.0500	0.0500	1.0000
Magnesium half sat. [mgMg/L]	0.1000	0.1000	1.0000
Cation half sat. [mmol/L]	0.1000	0.1000	1.0000
Calcium half sat. [mgCa/L]	0.1000	0.1000	1.0000
Aerobic/anoxic decay rate [1/d]	0.1000	0.1000	1.0000
Aerobic/anoxic maintenance rate [1/d]	0	0	1.0000
Anaerobic decay rate [1/d]	0.0400	0.0400	1.0000
Anaerobic maintenance rate [1/d]	0	0	1.0000
Sequestration rate [1/d]	4.5000	4.5000	1.0000
Anoxic growth factor [-]	0.3300	0.3300	1.0000

Print all Set current tab to default values OK Cancel

Delft Maintenance Approach Parameters

Model Change – Unified Hydrolysis Formulation

Different kinetic expressions were employed in BioWin 3.1 for the hydrolysis of particulate organic substrate mediated by activated sludge and anaerobic digestion. In Version 4.0, the kinetic expression for activated sludge hydrolysis is now also applied in anaerobic digesters. However, based on calibration, the *Anaerobic hydrolysis factor [AD]* applied in anaerobic digesters and active primary settlers is higher than the *Anaerobic hydrolysis factor [AS]* applied in anaerobic zones of activated sludge bioreactors.

The activated sludge anaerobic factor is lower than the default in BioWin 3.1; that value appeared to result in excessive VFA generation as a result of too much hydrolysis.

The unified hydrolysis formulation allows BioWin 4.0 users to effectively use an activated sludge element to model a non-conventional fermenter (such as a RAS fermenter) and obtain similar results as with an anaerobic digester element.

Name	Default	Value	Antonieus
Max. spec. growth rate [1/d]	3.2000	3.2000	1.0290
Substrate half sat. [mgCOD/L]	5.0000	5.0000	1.0000
Anoxic growth factor [-]	0.5000	0.5000	1.0000
Denitr N2 producers (NO3 or NO2) [-]	0.5000	0.5000	1.0000
Aerobic decay rate [1/d]	0.6200	0.6200	1.0290
Anoxic decay rate [1/d]	0.2330	0.2330	1.0290
Anaerobic decay rate [1/d]	0.1310	0.1310	1.0290
Hydrolysis rate [1/d]	2.1000	2.1000	1.0290
Hydrolysis half sat. [-]	0.0600	0.0600	1.0000
Anoxic hydrolysis factor (A) [-]	0.2800	0.2800	1.0000
Anaerobic hydrolysis factor (AS) [-]	0.0400	0.0400	1.0000
Anaerobic hydrolysis factor (AD) [-]	0.2000	0.2000	1.0000
Adsorption rate of colloids [L/(mgCOD d)]	0.1500	0.1500	1.0290
Ammonification rate [L/(mgN d)]	0.0400	0.0400	1.0290
Assimilative nitrate/nitrite reduction rate [1/d]	0.5000	0.5000	1.0000
Fermentation rate [1/d]	1.6000	1.6000	1.0290
Fermentation half sat. [mgCOD/L]	5.0000	5.0000	1.0000
Fermentation growth factor (AS) [-]	0.2500	0.2500	1.0000
Endogenous products decay rate [1/d]	0	0	1.0000
Free nitrous acid inhibition [mmol/L]	1.000E-7	1.000E-7	1.0000

Anaerobic Hydrolysis Factor Defaults

Model Changes – Parameter Default Changes

A few parameter default values in BioWin are changed in Version 4.0. Mostly these are very minor changes. However, attention should be drawn to two specific changes:

- Heterotroph biomass COD fraction in Influent elements;
- Bio-P model parameters (four values adjusted).

Influent Biomass

Traditionally it has been assumed that municipal influent wastewater contains a minimal amount of active biomass. Over the past three years EnviroSim has conducted extensive testing on influent wastewaters from many plants to quantify the heterotrophic biomass content. The Wentzel/UCT batch method has been applied where the oxygen utilization rate in a non-seeded influent wastewater sample is expected to increase exponentially over the first part of the test, starting from an initial low value (e.g. 1-2 mg/L/hour). Many of the EnviroSim experiments have indicated the active biomass content in the influent wastewater is appreciable. The plot below shows an example OUR response. The initial high OUR and the near-linear increase in OUR both indicate an appreciable amount of active heterotrophs in the wastewater.

The COD and BOD Influent element default COD fractions of heterotrophs (f_{zbh}) in BioWin 4.0 are changed to 0.02 to replace the previous value of 0.0001.

Editing COD Influent0

Input Type WW Fractions Monitor items

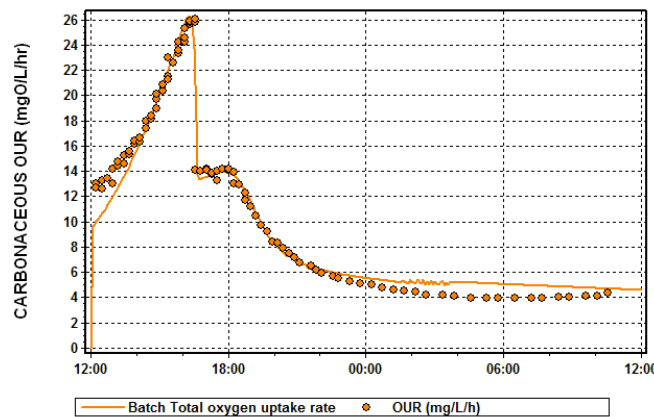
Name	Default	Value
Fbs - Readily biodegradable (including Acetate) [gCOD/g of total COD]	0.1600	0.1600
Fac - Acetate [gCOD/g of readily biodegradable COD]	0.1500	0.1500
Fxsp - Non-colloidal slowly biodegradable [gCOD/g of slowly degradable COD]	0.7500	0.7500
Fus - Unbiodegradable soluble [gCOD/g of total COD]	0.0500	0.0500
Fup - Unbiodegradable particulate [gCOD/g of total COD]	0.1300	0.1300
Fna - Ammonia [gNH3N/gTKN]	0.6800	0.6800
Fnox - Particulate organic nitrogen [gN/g Organic N]	0.5000	0.5000
Fnus - Soluble unbiodegradable TKN [gN/gTKN]	0.0200	0.0200
FupN - N:COD ratio for unbiodegradable part. COD [gN/gCOD]	0.0350	0.0350
Fpo4 - Phosphate [gPO4-P/gTP]	0.5000	0.5000
FupP - P:COD ratio for unbiodegradable part. COD [gP/gCOD]	0.0110	0.0110
Fzbh - OHO COD fraction [gCOD/g of total COD]	0.0200	0.0200
FZbm - Methyloph COD fraction [gCOD/g of total COD]	1.000E-4	1.000E-4
FZaob - AOB COD fraction [gCOD/g of total COD]	1.000E-4	1.000E-4
FZnob - NOB COD fraction [gCOD/g of total COD]	1.000E-4	1.000E-4
FZanob - ANAMMOX COD fraction [gCOD/g of total COD]	1.000E-4	1.000E-4

Set typical (Raw) Set typical (Settled)

Press F1 for help

OK Cancel

New default f_{zbh} (COD fraction of heterotrophs in influent) of 0.02



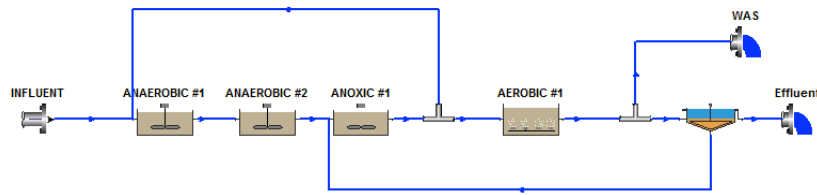
Example OUR response with default $f_{zbh} = 0.02$

Refined Bio-P Model Parameters

EnviroSim has conducted a thorough review of the biological P removal model predictions in light of other model changes such as the unified hydrolysis expression. The review was conducted by evaluating predictions of performance for many pilot-scale systems. As a result, default values for a few parameters have been adjusted slightly. The refined parameters include:

- Cation uptake
- Sequestration rate
- Aerobic P/ PHA ratio
- P/ Ac ratio

The predictive capacity of the model is demonstrated below for one of the pilot enhanced biological phosphorus removal (EBPR) systems (Wentzel's System 8a). This was a UCT configuration with recycle ratios of nitrified mixed liquor and clarifier underflow both at 100% of the influent flow. The SRT of the system was about 9 days.



The plots below show predictions for a range of parameters: VSS, OUR, phosphate and nitrate concentration profiles, and effluent TKN and COD. Good predictions of plant performance are achieved with the refined model parameters.

